

4. Dittrich AM, Meyer HA, Krokowski M, Quarcoo D, Ahrens B, Kube SM, Witzenrath M, Esworthy RS, Chu FF, and Hamelmann E. Glutathione peroxidase-2 protects from allergen-induced airway inflammation in mice. *Eur Respir J* 35: 1148–1154, 2010.
5. Esworthy RS, Aranda R, Martín MG, Doroshow JH, Binder SW, and Chu FF. Mice with combined disruption of Gpx1 and Gpx2 genes have colitis. *Am J Physiol Gastrointest Liver Physiol* 281: G848–G855, 2001.
6. Ho YS, Magnenat JL, Bronson RT, Cao J, Gargano M, Sugawara M, and Funk CD. Mice deficient in cellular glutathione peroxidase develop normally and show no increased sensitivity to hyperoxia. *J Biol Chem* 272: 16644–16651, 1997.
7. Hoffmann PR, Jourdan-Le Saux C, Hoffmann FW, Chang PS, Bollt O, He Q, Tam EK, and Berry MJ. A role for dietary selenium and selenoproteins in allergic airway inflammation. *J Immunol* 179: 3258–3267, 2007.
8. Sakamoto H, Imai H, and Nakagawa Y. Involvement of phospholipid hydroperoxide glutathione peroxidase in the modulation of prostaglandin D2 synthesis. *J Biol Chem* 275: 40028–40035, 2000.
9. Won HY, Sohn JH, Min HJ, Lee K, Woo HA, Ho YS, Park JW, Rhee SG, and Hwang ES. Glutathione peroxidase 1 deficiency attenuates allergen-induced airway inflammation by suppressing th2 and th17 cell development. *Antioxid Redox Signal* 13: 575–587, 2010.

Address correspondence to:
 Prof. Eckard Hamelmann
 University Children's Hospital
 Ruhr-University Bochum
 Bochum D-44791
 Germany

E-mail: e.hamelmann@klinikum-bochum.de

Abbreviations Used

GPx = glutathione peroxidases
 ROS = reactive oxygen species
 Th = T helper cell

DOI: 10.1089/ars.2010.3591.rs

Response to Meyer *et al.*

Eun Sook Hwang

To the Editor:

MEYER *ET AL.* ATTEMPTED to interpret the seemingly opposite effects of glutathione peroxidase (GPx) 1 and 2 in airway inflammation as reported in our (5) and their (2) works. Lung epithelial cells are important for the expression of a variety of chemokines in airway inflammation. In particular, eotaxin expression by lung epithelial cells increases eosinophil infiltration, which is a prominent characteristic of chronic inflammation. Although the specific expression of GPx2 in lung epithelial cells and its protective roles in ovalbumin-induced airway inflammation have been clearly validated (2), the molecular mechanisms have not yet been distinctly characterized. For instance, the effects of GPx2 (or reactive oxygen species [ROS]) on the expression of chemokines such as IP-10, MIG, CXCL10, and eotaxin in lung epithelial cells would be good to explain the protective roles of GPx2 in allergic airway inflammation. In addition, GPx2 suppresses COX2 expression in tumor migration and invasion (1), indicating that GPx2-mediated inhibition of COX-2 may prevent the synthesis of inflammation mediators leukotrienes, thromboxanes, and prostaglandins in lung epithelial cells. It is important to note that a recent report suggests that GPx1 specifically controls Th-cell differentiation by modulating ROS and subsequently inflammatory immune response (5). GPx1 deficiency increased ROS in T cells, thus

resulting in decreased Th2- and Th17-cell development and attenuated airway inflammation. GPx1 is thus suggested to play a key role in fine-tuning ROS level during Th-cell differentiation. However, the precise functions of GPx1 in lung epithelial cells remain to be clarified. In addition, GPx isoforms such as GPx2, GPx1, and GPx4 could be increased in the lung by antigenic challenge (3, 4). GPx2 expression is specifically increased by NRF2 activation, whereas GPx1 and GPx4 were prominently induced by selenium in a dose-dependent manner, implying the presence of distinct signaling pathway to induce GPx isoforms in a cell-specific and signal-dependent manner. It is therefore reasonable to conclude that both GPx1 and GPx2 eliminate ROS, but the expression and regulatory mechanisms of GPx1 and GPx2 may be distinct across cell types.

References

1. Banning A, Kipp A, Schmitmeier S, Lowinger M, Florian S, Krehl S, Thalmann S, Thierbach R, Steinberg P, and Brigelius-Flohe R. Glutathione peroxidase 2 inhibits cyclooxygenase-2-mediated migration and invasion of HT-29 adenocarcinoma cells but supports their growth as tumors in nude mice. *Cancer Res* 68: 9746–9753, 2008.
2. Dittrich AM, Meyer HA, Krokowski M, Quarcoo D, Ahrens B, Kube SM, Witzenrath M, Esworthy RS, Chu FF, and

- Hamelmann E. Glutathione peroxidase-2 protects from allergen-induced airway inflammation in mice. *Eur Respir J* 35: 1148–1154, 2010.
3. Romanowska M, Kikawa KD, Fields JR, Maciag A, North SL, Shiao YH, Kasprzak KS, and Anderson LM. Effects of selenium supplementation on expression of glutathione peroxidase isoforms in cultured human lung adenocarcinoma cell lines. *Lung Cancer* 55: 35–42, 2007.
 4. Singh A, Rangasamy T, Thimmulappa RK, Lee H, Osburn WO, Brigelius-Flohe R, Kensler TW, Yamamoto M, and Biswal S. Glutathione peroxidase 2, the major cigarette smoke-inducible isoform of GPX in lungs, is regulated by Nrf2. *Am J Respir Cell Mol Biol* 35: 639–650, 2006.
 5. Won HY, Sohn JH, Min HJ, Lee K, Woo HA, Ho YS, Park JW, Rhee SG, and Hwang ES. Glutathione peroxidase 1 deficiency attenuates allergen-induced airway inflammation by suppressing Th2 and Th17 cell development. *Antioxid Redox Signal* 13: 575–587, 2010.

Address correspondence to:
 Assoc. Prof. Eun Sook Hwang
 Science Building C206
 Ewha Womans University
 11-1 Daehyun-dong, Seodaemun-gu
 Seoul 120-750
 Korea

E-mail: eshwang@ewha.ac.kr

Date of first submission to ARS Central, August 24, 2010; date of final revised submission, September 12, 2010; date of acceptance, September 12, 2010.

This article has been cited by: